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Author for correspondence:

Benedikt R. Schmidt

e-mail: benedikt.schmidt@ieu.uzh.ch

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Dynamics of host populations affected by the emerging fungal pathogen *Batrachochytrium salamandrivorans*

Benedikt R. Schmidt^{1,2}, Claudio Bozzuto³, Stefan Lötters⁴ and Sebastian Steinfartz⁵

¹Institut für Evolutionsbiologie und Umweltwissenschaften, Universität Zürich, Winterthurerstrasse 190, 8057 Zürich, Switzerland

²karch, Passage Maximilien-de-Meuron 6, 2000 Neuchâtel, Switzerland

³Wildlife Analysis GmbH, Oetlisbergstrasse 38, 8053 Zürich, Switzerland

⁴Institut für Biogeographie, Universität Trier, Universitätsring 15, 54296 Trier, Germany

⁵Department of Evolutionary Biology, Technische Universität Braunschweig, Zoological Institute, Mendelssohnstrasse 4, 38106 Braunschweig, Germany

BRS, 0000-0002-4023-1001

Emerging infectious diseases cause extirpation of wildlife populations. We use an epidemiological model to explore the effects of a recently emerged disease caused by the salamander-killing chytrid fungus *Batrachochytrium salamandrivorans* (*Bsal*) on host populations, and to evaluate which mitigation measures are most likely to succeed. As individuals do not recover from *Bsal*, we used a model with the states susceptible, latent and infectious, and parametrized the model using data on host and pathogen taken from the literature and expert opinion. The model suggested that disease outbreaks can occur at very low host densities (one female per hectare). This density is far lower than host densities in the wild. Therefore, all naturally occurring populations are at risk. *Bsal* can lead to the local extirpation of the host population within a few months. Disease outbreaks are likely to fade out quickly. A spatial variant of the model showed that the pathogen could potentially spread rapidly. As disease mitigation during outbreaks is unlikely to be successful, control efforts should focus on preventing disease emergence and transmission between populations. Thus, this emerging wildlife disease is best controlled through prevention rather than subsequent actions.

1. Introduction

Emerging infectious diseases threaten wildlife populations because they can cause mass mortality, which may ultimately lead to local and global extinction of hosts. Such extinctions may cause the loss of evolutionary diversity and can lead to changes in ecosystem function [1–4]. Emerging infectious diseases pose a major challenge to conservation biologists and practitioners because the effects of emerging pathogens, often new to science, on host populations are very strong and because disease mitigation in wildlife populations is still in its infancy [5,6]. Appropriate management actions are currently only available for the pre-invasion stage of pathogen emergence (i.e. biosecurity) whereas methods for pathogen control and mitigation during and after pathogen emergence have not yet been developed [7], but there are some proof-of-concept studies showing that mitigation is possible in the wild [8,9]. Mathematical models have proven utility in understanding disease dynamics and exploring management strategies [10]. Here, we use an epidemiological model to investigate the dynamics of a fungal disease in salamanders, caused by *Batrachochytrium salamandrivorans* ('the devourer of salamanders', hereafter *Bsal*), that recently emerged in Europe [11–14], with the goal of informing mitigation strategies.

Standard epidemiological theory suggests that pathogens are unlikely to drive hosts to extinction [15]. Emerging fungal pathogens, however, can, particularly if host populations are large [4]. Emerging fungal or fungal-like diseases have led already to mass mortality, local population extirpations and regional extinction of various hosts, including soft corals, bees, bats, frogs, salamanders and snakes [4]. Like many other emerging wildlife diseases [16], *Bsal* was most probably brought from Asia to Europe through the animal trade. *Bsal* emerged in The Netherlands in wild salamander populations, where it caused mass mortality and drove salamander populations to the edge of extirpation [11–13]. This novel pathogen is of global conservation concern because it could have devastating effects on salamander biodiversity worldwide, as well as knock-on consequences for ecosystem function [17,18]. While there is consensus that preventing the invasion of *Bsal* into new areas should be a priority [17,18], little is known about intervention strategies once *Bsal* has emerged in an area. Upon detection in a new locality, immediate management actions to prevent the spread of the pathogen, such as restricting site-level access, decontaminating a site and removal of amphibians from the site, should be considered [18]. Because uncertainties hinder the effective deployment of interventions in areas where *Bsal* has emerged [18], we modelled the temporal and spatial dynamics of the *Bsal*–salamander system, with the ultimate goal of informing mitigation strategies.

2. Model

To gauge the ecological consequences of a *Bsal* emergence and spread, we considered two scenarios. (i) What happens to a host population if *Bsal* emerges locally? (ii) How does *Bsal* spread over an extended area? In the latter scenario, we were also interested in the effects that human-mediated dispersal of infected salamanders have on the overall spread of *Bsal*. Since the spread of *Bsal* is important in our analysis, we considered model formulations reflecting both temporal and spatio-temporal changes.

2.1. Basic model: temporal formulation

We used an epidemiological dynamic model, which differed from previous amphibian–chytrid models where the size of the aquatic zoospore population was modelled (i.e. an index of pathogen abundance [19,20]). Currently, we lack information on *Bsal* zoospores. Since infection of salamander larvae by *Bsal* is not yet reported, we modelled the adult female portion of a host population by considering susceptible individuals, latent individuals (infected but not infectious yet) and infectious individuals. The basic model is given by the following system of equations:

$$\left. \begin{aligned} \frac{dS}{dt} &= (b - d)S - \gamma SN - \beta SI, \\ \frac{dL}{dt} &= \beta SI - (d + e)L - \gamma LN \\ \text{and} \quad \frac{dI}{dt} &= eL - (d + d_1)I - \gamma IN. \end{aligned} \right\} \quad (2.1)$$

Susceptible individuals (S) produce offspring at a rate b and die at a rate d . As we focus on the adult (female) portion of a population, b is the rate at which adults are recruited per unit time (1 year). To these

rates we add a density-dependent reduction in population growth [21] $-\gamma SN$, by assuming that the *per capita* effect of density dependence is a function of all adults, i.e. $N = S + L + I$. Note that γ is related to carrying capacity $K = (b - d)/\gamma$. For all analyses, we use densities (individuals per hectare). Finally, the growth rate of susceptible individuals is also diminished by contact with infected individuals, i.e. βSI (representing true mass action, *sensu* [22]). New individuals enter the latent portion of the population (L) as infected, but not yet infectious, individuals. We assume that their contribution to reproduction is negligible. Thus, latent individuals either die naturally, $-(dL + \gamma LN)$, or become infectious at a rate e . These newly generated infectious individuals (I) are also assumed not to contribute to reproduction, and either die naturally, $-(dI + \gamma IN)$, or because of *Bsal* ($d_I I$). An identical model, resulting from analysis of the spread of rabies [23], is summarized and discussed by, among others, Shigesada & Kawasaki [24].

Immediately before *Bsal* enters a population, the population (composed only of S) is assumed to be at K . The condition for *Bsal* to spread is given by the following equation (see [24] for a derivation):

$$K > \frac{(b + e)(b + d_I)}{\beta e} = K_{\text{threshold}}. \quad (2.2)$$

Thus, if K is higher than $K_{\text{threshold}}$, *Bsal* will spread; otherwise it will die out. This threshold can also be used to calculate the proportion of the host population to be removed to prevent a disease outbreak, $p_{\text{remove}} = 1 - K_{\text{threshold}}/K$ [23]. If removal is not feasible, in terms of either effort or public acceptance, then another way of thinking about how to prevent a disease outbreak is to ask what mitigation actions could influence model parameters in equation (2.2) such that $K_{\text{threshold}}$ becomes higher than the actual K . A good approximation in the present case is $d_I/\beta \geq K$; for more details, see the electronic supplementary material.

If *Bsal* spreads, equation (2.1) has a non-trivial equilibrium given by the following equation:

$$\left. \begin{aligned} S_{\text{eq}} &= K - L_{\text{eq}} - I_{\text{eq}} - \frac{\beta K I_{\text{eq}}}{(b - d)}, \\ L_{\text{eq}} &= \frac{(b + d_I - \beta I_{\text{eq}}) I_{\text{eq}}}{e}, \\ I_{\text{eq}} &= \frac{(b - d)(K\beta e - (b + e)(b + d_I))}{\beta(K\beta e - b(b - d))}. \end{aligned} \right\} \quad (2.3)$$

and

2.2. Spatio-temporal formulation

To analyse the spatial spread of *Bsal*, we expanded our basic model equation (2.1) by including diffusion terms. We assumed that diffusion does not differ between dimensions, leading to a concentric spread of *Bsal* in homogeneous space. We thus modelled diffusion in one spatial dimension (x). We added to every equation a (spatial) diffusion term $D_j(\partial^2 j/\partial x^2)$, where $j = S, L$ or I . While we note that real landscapes are heterogeneous for both host and pathogen [25,26], we are lacking information on the diffusion ability of the different members of the population (S, L, I). Thus, we assumed $D_S = D_L = D_I = D$. As before, we calculated conditions for *Bsal* to spread, starting with the situation that right before *Bsal* enters the population, the population (all S) is at K . Assuming a homogeneous environment, this condition is identical to equation (2.2), i.e. for the basic model without diffusion [24]. Shigesada & Kawasaki [24] showed that if the conditions for *Bsal* to spread are met, then the range of expansion will behave like a propagating wave. The constant speed c of the wavefront is given by the following equation [24]:

$$c = \sqrt{2D \left(\sqrt{(e - d_I)^2 + 4e\beta K} - 2b - d_I - e \right)}. \quad (2.4)$$

Finally, we analysed how *Bsal* would spread over an extended area assuming that, in addition to the local spread velocity given by equation (2.4), a small number of infected salamanders is unintentionally and randomly released by humans into healthy populations elsewhere, or that a vector (e.g. wildfowl [27]) transports the pathogen to new sites. The situation that we envisaged can be emulated with a 'scattered colony model' [28]: (i) *Bsal* spreads locally and (ii) introduced infectious individuals in healthy populations show a sufficiently large distance to other colonies, so that over a restricted period of time the local *Bsal* spread does not lead to overlapping infected colonies. One main result from such a modelling approach states how the total area of all infected colonies changes with time (for a derivation, see [28]):

$$A(t) = \frac{2\pi c^2}{\mu} \left(\frac{e^{\mu t} - 1}{\mu} - t \right), \quad (2.5)$$

Table 1. Parameter descriptions and values used in this study.

parameter	description	value	source
b	birth rate: number of new adult females per adult female	0.39 yr^{-1}	[29]
d	death rate: $1/d =$ life expectancy of an adult	$1/d \approx 8 \text{ yr}$, $d \approx 0.125 \text{ yr}^{-1}$	[29]: $1/d$
d_i	death rate infected salamanders: $1/d_i =$ life expectancy of infected salamander	$1/d_i \approx 7 \text{ days}$, $d_i = 52 \text{ yr}^{-1}$	[11]: $1/d_i$
γ	strength of density dependence: related to carrying capacity $K = (b - d)/\gamma$	$K = 25$ females/hectare (ha); compared to densities in table 7.3 in [29], this density is at the lower end of estimated ones. Higher densities would lead to even worse model results	authors' estimate, based on [29]
β	transmission coefficient: $1/\beta =$ average time period before an infected salamander encounters another salamander, assuming successful infection given an encounter	$1/\beta \approx 1 \text{ week}$, $\beta = 52 \text{ ha yr}^{-1}$	authors' estimate
e	rate at which latents become infectious; $1/e =$ average latent period	$1/e \approx 1 \text{ day}$, $e = 364 \text{ yr}^{-1}$	[11]: $1/e$
D_j	diffusion coefficient of $j = S, L$ or I ; $D_j = \langle x^2 \rangle_j / (4t)$; t is time in years and $\langle x^2 \rangle$ is the mean square of the straight line that a salamander travels in a year	$\chi \approx 0.5 \text{ km yr}^{-1}$	authors' estimate
μ	colonization rate	$\mu \approx 1 \text{ yr}^{-1}$	authors' estimate

where μ is a fixed rate at which infected individuals originating from an infected colony are brought into healthy colonies per year. To make the results from equation (2.5) more tangible, we related it proportionally to the total suitable habitat for salamanders in Germany; for more details, see the electronic supplementary material.

2.3. Parameter values

To date, *Bsal* mainly affects populations of the fire salamander, *Salamandra salamandra* [14]. Therefore, the best data are available for this host species. Therefore, we parametrized the model with data from this host species (table 1).

To gauge the effect of parameters on the results, we performed differential sensitivity analyses, with the assumption that parameters are uncorrelated. Furthermore, we show results for the effect of a crucial parameter in our study, K , by numerically solving equation (2.1). To this end, we assumed an initial density of one infectious individual per square kilometre entering a healthy population.

3. Results

The threshold population density for *Bsal* to spread (equation (2.2)) is $K_{\text{threshold}} = 1.0086$ adult females ha^{-1} ; that is, approximately 4% of K . Furthermore, equilibrium density of susceptibles is $S_{\text{eq}} = 1.0027$ adult females ha^{-1} . If the condition given by equation (2.2) holds, then this equilibrium is either stable or undergoes a Hopf bifurcation at $K_{\text{H}} \approx 9.5$, meaning that, if $K > K_{\text{H}}$, the system is characterized by a stable limit cycle, oscillating about S_{eq} . To pre-emptively counteract a disease outbreak, the percentage of adults to be removed should be at least $p_{\text{remove}} \approx 96\%$. Electronic supplementary material, figure S1a,b shows the sensitivity of $K_{\text{threshold}}$ to proportional changes in parameter values, either in absolute terms (electronic supplementary material, figure S1a), or as deviations from $K_{\text{threshold}}$, calculated using point-estimate

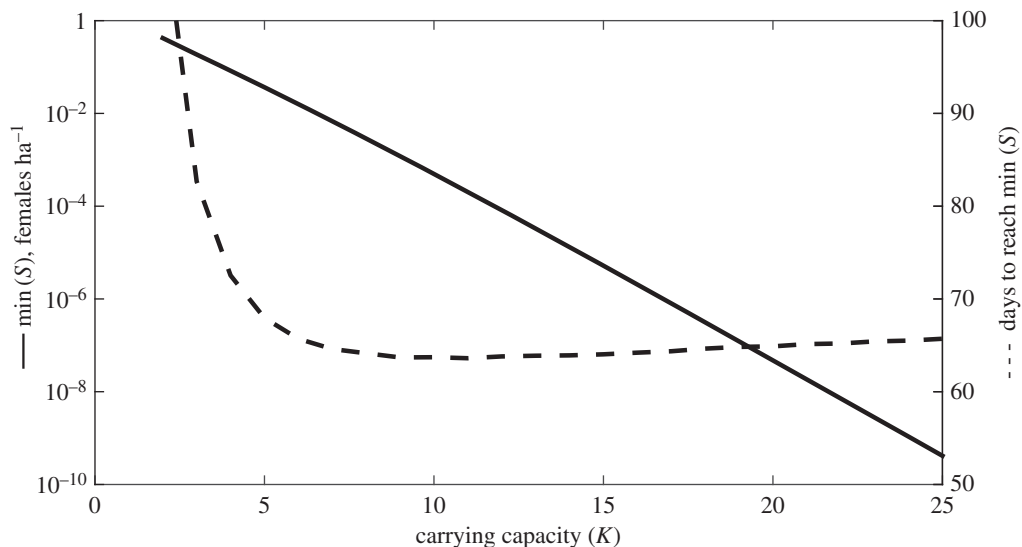


Figure 1. The effect of varying carrying capacity K on the minimal density reached by S ($\min(S)$, left y-axis) and time for S to reach $\min(S)$ (right y-axis).

parameter values (in %, electronic supplementary material, figure S1*b*). As is evident, even varying single parameters by 20% leads to an increase of $K_{\text{threshold}}$ of at most 25%.

In the case of a disease outbreak, for realistic values of K the trajectory connecting K at t_0 to the equilibrium state reaches densities of almost zero adult females ha^{-1} (figure 1). The figure also shows the time needed for S to reach its minimal density, that is, the minimal density during the ‘population crash’; since we had to rely on the literature and expert opinion to parametrize our model, we gauged this approach to be more appropriate than using a threshold density. This minimum seems to be reached within two to three months.

By expanding the basic model to include spatial diffusion, the calculated constant speed of the propagation wave is $c = 11.15 \text{ km yr}^{-1}$. Electronic supplementary material, figure S1*c,d* shows its sensitivity to proportional parameter changes. Yachi *et al.* [30] showed for the fox–rabies system that—assuming a constant homogeneous environment like in the present study—the conditions for spatial propagation to occur are given by equation (2.2). Further, these authors also analysed how the propagation wave develops. Here, we wish to highlight one result: if $K > K_H$ (i.e. where a stable limit cycle governs the local dynamics), the propagation wave is characterized by violent and irregular fluctuations before S_{eq} is reached.

Electronic supplementary material, figure S3 shows the proportion of suitable habitat that becomes infected as a function of time, and a fixed rate μ ; the sensitivity to parameters is shown in the electronic supplementary material, figure S1*ef*. Our educated guess with regard to the total suitable habitat for *Salamandra salamandra* in Germany is that it amounts to around 258 750 km^2 (based on Sillero *et al.* [31]). For the sake of readability, we show two fixed rates: 1 yr^{-1} and 0.1 yr^{-1} . After 8 years with the first rate, approximately 9% of all suitable habitats would be infected, whereas the latter rate would result in approximately 0.1%. Nonetheless, note the exponential nature of equation (2.5).

4. Discussion

Epidemiologists have long used mathematical models to better understand the temporal and spatial dynamics of disease in host populations. Owing to the recent nature of *Bsal* emergence, we lack spatio-temporal *Bsal*-related epidemiological data and must rely on the limited information available to estimate model parameters. However, in general, our model is little affected by uncertainty in parameter estimates (electronic supplementary material, figure S1). The discussion will therefore focus on control of the disease.

Our model confirms field-based observations that the fungus can lead to the collapse of host populations within months or even weeks [13], and suggests that *Bsal* can spread rapidly. Our model suggests a speed of approximately 11 km yr^{-1} , which is comparable (by order of magnitude) to estimates

for the closely related pathogenic amphibian chytrid *Batrachochytrium dendrobatidis* [32]. Recent work suggests that *Bsal* is more widely distributed than previously thought in wild European salamander populations [14]. This may be the result of dispersal from the index site. If so, then the available surveillance data [14] suggest that the estimate of approximately 11 km yr^{-1} may be an overestimate. Alternatively, the spatial distribution of *Bsal* may have remained unchanged, and the increase of known occurrences is the result of improved surveillance.

If the host population is below a threshold size, disease outbreaks are unlikely and an epizootic will fade out once host population size has been strongly reduced [15]. The general host–fungal pathogen model of Fisher *et al.* [4] proposes that there may be no threshold population size that prevents outbreaks of fungal diseases, and that strong disease-induced population declines are the likely consequence. For *Bsal*, there also seems to be no biologically meaningful threshold of host population size that may prevent an outbreak. The outbreak threshold (equation (2.2)) is a small fraction of what we assume to be a low-density population, suggesting that *Bsal* poses a risk for all salamander populations. Our results suggest that removal or culling is unlikely to work in practice, whereas it may be possible for the closely related chytrid fungus *Batrachochytrium dendrobatidis* [33]. Other control strategies are also likely to fail because the parameters that determine the outbreak threshold would have to be strongly altered (electronic supplementary material, figure S2).

Our model predicts that an outbreak of *Bsal* is likely to cause a rapid collapse of the host population (figure 1). Although, deterministically, an equilibrium state will eventually be attained, it is unlikely that a real population will survive such an extreme bottleneck. We conclude that an outbreak should fade out quickly as the host population is rapidly depleted. Additionally, because *Bsal* is most likely transmitted by direct contact between adult individuals, transmission becomes unlikely at densities as low as the estimated $K_{\text{threshold}}$. Reservoir hosts might change transmission dynamics and the epidemiology of *Bsal* with likely consequences for disease control [15,34]. Yet, while it is known that *Bsal* is a multihost pathogen [10], there are no published studies that describe epidemiologically relevant reservoir hosts. This has three major implications for disease control and spread. First, if there is an outbreak in a population, mitigation is unlikely to succeed during the outbreak. Second, the pathogen is unlikely to persist in the absence of an environmental reservoir or reservoir host. Third, because salamanders do not move much in a matter of months [25,35], it seems unlikely that infected individuals would move far enough to transmit the disease to nearby neighbouring forest patches with salamander demes where conditions are suitable for *Bsal* [25,26]. Our spatial model may therefore overestimate the spread of the pathogen.

Bsal is a newly emerged infectious disease that threatens salamander and newt biodiversity in Europe. Our model predicts, and thereby confirms empirical results, that *Bsal* can have strong negative effects on host populations. Mitigating the effects of this disease is a conservation priority. As outbreaks are unlikely to be controllable, the focus should be on limiting pathogen spread among sites and populations and limiting establishment at new sites [7]. Our model predicts that the pathogen will spread at a rate of approximately 11 km yr^{-1} , if local populations do not become extinct. Controlling spatial spread is therefore a formidable task because we do not know yet how *Bsal* spreads spatially. Therefore, studying the spatial epidemiology of *Bsal* should become a research priority.

If limiting the spread of *Bsal* is the best way to control this emerging pathogen, then it is necessary to avoid human-mediated spread, as it may occur through the pet trade [36]. Human-mediated spread might greatly facilitate the spread of *Bsal* (electronic supplementary material, figure S3). It is therefore important to enforce biosafety rules for biologists conducting fieldwork on amphibians, and to inform herpetologists, naturalists and captive breeders that salamanders must not be translocated. Thus, as is the case for emerging wildlife diseases in general, prevention of emergence and spread is more effective than responses at later stages of the invasion [7,33].

Data accessibility. This paper contains no data. The sources of the data that we used to parametrize the model are listed in table 1 and in the list of references [11,29].

Authors' contributions. B.R.S. wrote the paper with input from all authors. C.B. conceived the study and performed all model-related work and analyses and contributed to writing. S.L. contributed to writing. S.S. conceived the study and contributed to writing. All authors gave their final approval for publication.

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References

- Daszak P, Cunningham AA, Hyatt AD. 2000 Emerging infectious diseases of wildlife—threats to biodiversity and human health. *Science* **287**, 443–449. (doi:10.1126/science.287.5452.443)
- Colón-Gaud C, Whiles MR, Kilham SS, Lips KR, Pringle CM, Connelly S, Peterson SD. 2009 Assessing ecological responses to catastrophic amphibian declines: patterns of macroinvertebrate production and food web structure in upland Panamanian streams. *Limnol. Oceanogr.* **54**, 331–343. (doi:10.4319/lo.2009.54.1.0331)
- Crawford AJ, Lips KR, Bermingham E. 2010 Epidemic disease decimates amphibian abundance, species diversity, and evolutionary history in the highlands of central Panama. *Proc. Natl Acad. Sci. USA* **107**, 13 777–13 782. (doi:10.1073/pnas.0914115107)
- Fisher MC, Henk DA, Briggs CJ, Brownstein JS, Madoff LC, McCraw SL, Gurr SJ. 2012 Emerging fungal threats to animal, plant and ecosystem health. *Nature* **484**, 186–194. (doi:10.1038/nature10947)
- Woodhams DC *et al.* 2011 Mitigating amphibian disease: strategies to maintain wild populations and control chytridiomycosis. *Front. Zool.* **8**, 8. (doi:10.1186/1742-9994-8-8)
- Scheele BC, Hunter DA, Grogan LF, Berger LE, Kolby JE, McFadden MS, Marantelli G, Skerratt LF, Driscoll DA. 2014 Interventions for reducing extinction risk in chytridiomycosis-threatened amphibians. *Conserv. Biol.* **28**, 1195–1205. (doi:10.1111/cobi.12322)
- Langwig KE *et al.* 2015 Context-dependent conservation responses to emerging wildlife diseases. *Front. Ecol. Environ.* **13**, 195–202. (doi:10.1890/140241)
- Bosch J, Sanchez-Tomé E, Fernández-Loras A, Oliver JA, Fisher MC, Garner TWJ. 2015 Successful elimination of a lethal wildlife infectious disease in nature. *Biol. Lett.* **11**, 20150874. (doi:10.1098/rsbl.2015.0874)
- Hudson MA *et al.* 2016 In-situ Itraconazole treatment improves survival rate during an amphibian chytridiomycosis epidemic. *Biol. Conserv.* **195**, 37–45. (doi:10.1016/j.biocon.2015.12.041)
- Ferguson NM, Cummings DA, Fraser C, Cajka JC, Cooley PC, Burke DS. 2006 Strategies for mitigating an influenza pandemic. *Nature* **442**, 448–452. (doi:10.1098/rsif.2009.0312)
- Martel A *et al.* 2013 *Batrachochytrium salamandrivorans* sp. nov. causes lethal chytridiomycosis in amphibians. *Proc. Natl Acad. Sci. USA* **110**, 15 325–15 329. (doi:10.1073/pnas.1307 356110)
- Martel A *et al.* 2014 Recent introduction of a chytrid fungus endangers Western Palearctic salamanders. *Science* **346**, 630–631. (doi:10.1126/science.1258268)
- Spitzen-van der Sluijs A, Spikmans F, Bosman W, de Zeeuw M, van der Meij T, Govers E, Kik M, Pasmans F, Martel A. 2013 Rapid enigmatic decline drives the fire salamander (*Salamandra salamandra*) to the edge of extinction in the Netherlands. *Amphib.-Reptil.* **34**, 233–239. (doi:10.1163/15685381-00002891)
- Spitzen-van der Sluijs A *et al.* 2016 Expanding distribution of lethal amphibian fungus *Batrachochytrium salamandrivorans* in Europe. *Emerg. Infect. Dis.* **22**, 1286–1288. (doi:10.3201/eid2207.160109)
- De Castro F, Bolker B. 2005 Mechanisms of disease-induced extinction. *Ecol. Lett.* **8**, 117–126. (doi:10.1111/j.1461-0248.2004.00693.x)
- Dunn AM, Hatcher MJ. 2015 Parasites and biological invasions: parallels, interactions, and control. *Trends Parasitol.* **31**, 189–199. (doi:10.1016/j.pt.2014.12.003)
- Gray MJ *et al.* 2015 *Batrachochytrium salamandrivorans*: the North American response and a call for action. *PLoS Pathog.* **11**, e1005251. (doi:10.1371/journal.ppat.1005251)
- Grant EHC *et al.* 2016 Salamander chytrid fungus (*Batrachochytrium salamandrivorans*) in the United States—developing research, monitoring, and management strategies. US Geological Survey Open-File Report 2015-1233. (doi:10.3133/ofr20151233)
- Mitchell KM, Churcher TS, Garner TW, Fisher MC. 2008 Persistence of the emerging pathogen *Batrachochytrium dendrobatidis* outside the amphibian host greatly increases the probability of host extinction. *Proc. R. Soc. B* **275**, 329–334. (doi:10.1098/rspb.2007.1356)
- Briggs CJ, Knapp RA, Vredenburg VT. 2010 Enzootic and epizootic dynamics of the chytrid fungal pathogen of amphibians. *Proc. Natl Acad. Sci. USA* **107**, 9695–9700. (doi:10.1073/pnas.0912886107)
- Salvidio S. 2009 Detecting amphibian population cycles: the importance of appropriate statistical analyses. *Biol. Conserv.* **142**, 455–461. (doi:10.1016/j.biocon.2008.10.035)
- McCallum H, Barlow N, Hone J. 2001 How should pathogen transmission be modelled? *Trends Ecol. Evol.* **16**, 295–300. (doi:10.1016/s0169-5347(01) 02144-9)
- Anderson RM, Jackson HC, May RM, Smith AM. 1981 Population dynamics of fox rabies in Europe. *Nature* **289**, 756–771. (doi:10.1038/289765a0)
- Shigesada N, Kawasaki K. 1997 *Biological invasions: theory and practice*. Oxford, UK: Oxford University Press.
- Ficetola GF, Manenti R, De Bernardo F, Padoa-Schioppa E. 2012 Can patterns of spatial autocorrelation reveal population processes? An analysis with the fire salamander. *Ecography* **35**, 693–703. (doi:10.1111/j.1600-0587.2011. 06483.x)
- Feldmeier S, Schefczyk L, Wagner N, Heinemann G, Veith M, Lötters S. 2016 Exploring the distribution of the spreading lethal salamander chytrid fungus in its invasive range in Europe—a macroecological approach. *PLoS ONE* **11**, e0165682. (doi:10.1371/ journal.pone.0165682)
- Yachi S, Kawasaki K, Shigesada N, Teramoto E. 1989 Spatial patterns of propagating waves of fox rabies. *Forma* **4**, 3–12.
- Van Rooij P, Martel A, Haesebrouck F, Pasmans F. 2015 Amphibian chytridiomycosis: a review with focus on fungus-host interactions. *Vet. Res.* **46**, 137. (doi:10.1186/s13567-015-0266-0)
- Thiesmeier B. 2004 *Der Feuersalamander*. Bielefeld, Germany: Laurenti.
- Shigesada N, Kawasaki K, Takeda Y. 1995 Modeling stratified diffusion in biological invasions. *Am. Nat.* **1**, 229–251. (doi:10.1086/285796)
- Sillero N *et al.* 2014 Updated distribution and biogeography of amphibians and reptiles of Europe. *Amphib.-Reptil.* **35**, 1–31. (doi:10.1163/15685381- 00002935)
- Lips KR, Diffendorfer J, Mendelson JR III, Sears MW. 2008 Riding the wave: reconciling the roles of disease and climate change in amphibian declines. *PLoS Biol.* **6**, e72. (doi:10.1371/journal.pbio. 0060072)
- Garner TWJ, Schmidt BR, Martel A, Pasmans F, Muths E, Cunningham AA, Weldon C, Fisher MC, Bosch J. 2016 Mitigating amphibian chytridiomycoses in nature. *Phil. Trans. R. Soc. B* **371**, 20160207. (doi:10.1098/rstb.2016.0207)
- Hudson PJ, Rizzoli A, Grenfell BT, Heesterbeek H, Dobson AP (eds). 2001 *The ecology of wildlife diseases*. Oxford, UK: Oxford University Press.
- Schulte U, Küsters D, Steinfartz S. 2007 A PIT tag based analysis of annual movement patterns of adult fire salamanders (*Salamandra salamandra*) in a Middle European habitat. *Amphib.-Reptil.* **28**, 531–536. (doi:10.1163/156853807782152543)
- Cunningham AA *et al.* 2015 Emerging disease in UK amphibians. *Vet. Rec.* **176**, 468. (doi:10.1136/vr. h2264)